

Curriculum vitae for Jason R. Dwyer

Education	2
Research Experience	2
Significant Awards and Fellowships	3
Named Fellowships.....	5
Publications.....	5
Intellectual Property.....	8
Lectures and Presentations.....	8
Grant Funding	15
Trainee Supervision	16
Teaching Experience.....	18
Service.....	19
University of Rhode Island	19
Department of Chemistry.....	20
Professional (External) Service.....	22
Public Outreach and Media.....	23
Entrepreneurship & Consulting	25
Special Training	25
Professional Memberships	25

Jason R. Dwyer
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Education

- 1998-2005 **Ph.D., Physical Chemistry – Natural Sciences and Engineering Council of Canada Postgraduate Fellowships A&B**
Professor R.J. Dwayne Miller, Canada Research Chair in Femtoscience
Departments of Chemistry & Physics, University of Toronto, Toronto, Canada
Thesis: *Femtosecond Electron Diffraction Studies of Ultrafast Structural Dynamics*
-Designed, built and used a femtosecond electron diffractometer to probe ultrafast laser-driven structural dynamics. First observation of real-space structure with femtosecond temporal resolution. Designed nanofabricated tools to enhance instrument performance.
- 1994-1998 **B.Sc., Honours Chemistry and Full-Time Chemistry Research – Canada Scholar**
5/1998- Professor Stuart M. Rothstein
8/1998 Departments of Chemistry & Physics, Brock University, St. Catharines, Canada
Thesis: *Variational Monte-Carlo study of a core-valence partitioned wavefunction: Electron affinities for selected first-row atoms using a modest basis set.*
-Wrote custom variational Monte Carlo computer code to compute atomic properties and explore core-valence electron partitioning using explicitly correlated wavefunctions.

Research Experience

- 7/2020- **Professor of Chemistry**
Department of Chemistry, University of Rhode Island
- 7/2015- **Associate Professor of Chemistry**
6/2020 Department of Chemistry, University of Rhode Island
- 8/2009- **Assistant Professor of Chemistry**
6/2015 Department of Chemistry, University of Rhode Island
- Research:** Design, fabrication and use of single-molecule nanopore sensors for bioanalytical applications and biophysical investigations. Thin film surface chemical modification strategies to control interfacial properties in confined environments. Development and application of techniques for surface-enhanced Raman spectroscopy (SERS).
- Notable Themes:** (1) Developing nanopores as a tool for glycomics and industry-ready pharmaceutical quality assurance, (2) developing routes to chemically optimize nanopores for fundamental research and commercial applications spanning genomics, proteomics, and glycomics, and (3) fundamental and applied optical sensing research to support real-time environmental monitoring from ions such as nitrate to the scale of microbe.
- 2009-2014 **Co-Founder and Consulting Vice President of Research and Development, Insight Nanofluidics, Inc., Toronto, Canada**
-Led the grant-funding and technical development for commercialization of a nanofluidic sample cell for high-performance spectroscopy and transmission electron microscopy on liquid samples, including design and interpretation of proof-of-principle experiments.

02/2008-08/2009	Postdoctoral Research Fellow Professor Andre Marziali, Director of Engineering Physics Dept. of Physics & Astronomy, University of British Columbia, Vancouver, Canada -Built on expertise in chemistry, nanofabrication and project management to work towards design and deployment of a clinic-ready lab-on-a-chip genotyping device.
5/2005-12/2007	Postdoctoral Research – Natural Sciences and Engineering Research Council of Canada (NSERC) Postdoctoral Research Fellow Professor Dr. Thomas Elsaesser, Director, Dr. Erik T.J. Nibbering, Project Leader Max Born Institute for Nonlinear Optics and Short Pulse Spectroscopy, Berlin, Germany -Used femtosecond infrared spectroscopy and materials science to study DNA base pair hydrogen bonding and DNA hydration.
summers 1995-1997	Chemistry Research Assistant Professor Kathleen M. Gough Department of Chemistry, University of Manitoba, Winnipeg, Canada -Used commercial <i>ab initio</i> molecular modelling packages and wrote custom analysis software to investigate the effect of molecular conformation on the electronic structure of hydrocarbons, as manifested in their Raman spectra.

Significant Awards and Fellowships

Year	Award	Scope of Competition
	2019 FACSS Innovation Award Winner <i>[International conference established 1972, convened by a federation of 15 independent societies of analytical chemistry and spectroscopy, with strong academic and industrial representation. Four finalists were chosen from a competitive review of conference abstracts: “These awards are currently given for the most innovative and outstanding new research advancements debuted orally at the SciX Conference... Finalists are selected for presentations by a panel of judges and all finalist present their work at a special plenary session at the SciX conference. Award winners are selected by the judging panel based on the quality of the work presented, responses provided to the judges' questions after the plenary presentation, and responses to audience questions.”]</i>	
2019		International
2019	Nominee, Sheila Black Grubman Faculty Outstanding Service Award	URI
2018	Graduate Mentoring Excellence Award Recipient, Arts & Sciences Best Presentation, American Chemical Society Meeting, SESSION: Colloidal Metal & Semiconductor Nanostructures: Theory, Synthesis & Application. (<i>Thin-film nanofluidics for single-particle analysis</i>, paper # COLL 121; Session Chair Jing Zhao, University of Connecticut)	URI
2017	Canada 150 Research Chair Nominee (Department of Chemistry, Brock University)	254th ACS National Meeting
2017	<i>[The Canada 150 Research Chairs Program, funded at \$350k or \$1M/year for 7 years to attract top-tier, internationally based scholars and researchers to Canada.]</i>	International
2017	URI Intellectual Property Recognition Award (patent submission)	U. Rhode Island
2016	Extraordinary Salary Increase (ESI) <i>[Interviewed for Tier 1 Canada Research Chair (\$200k/year for 7 year term, renewable once) at Guelph University]</i>	U. Rhode Island/International
2015	URI Intellectual Property Recognition Award	U. Rhode Island
2014	URI Intellectual Property Recognition Award (patent submission+ invention disclosure)	U. Rhode Island

2013	Early Career Faculty Research Excellence Award: Life Sciences, Physical Sciences and Engineering, URI Division of Research and Economic Development	U. Rhode Island
2012	Extraordinary Salary Increase (ESI) <i>[for receiving an NSF CAREER award (on first application)]</i>	U. Rhode Island/National
2006	Top 5 finish in the Mathematics, Physical Sciences and Engineering division of the 2006 CGS/UMI Distinguished Dissertation Award competition.	North America
2006	Nomination finalist for the Governor General's Gold Medal Award <i>[The gold Governor General's Academic Medal is awarded for top performance at the graduate level at a Canadian educational institution.]</i>	U. of Toronto/National
2005-2007	Natural Sciences and Engineering Research Council of Canada (NSERC) Postdoctoral Fellowship (\$80,000 total). <i>[The Canadian equivalent of the National Science foundation.]</i>	Canada
2005	Larry Calvert Travel Award to IUCr2005 (International Union for Crystallography Annual Meeting, \$1,500)	Canada
2005	IUCr2005 Travel Bursary (August 2005; Florence, Italy)	International
2004-2005	Donald J. Le Roy Prize in Physical Chemistry (\$500 + award lecture) <i>[the top research prize in graduate physical chemistry at UofT]</i>	U. of Toronto
2003	Michael J. Dignam Graduate Travel Award (CLEO Pacific Rim, December 2003; Taipei, Taiwan, \$1,000)	U. of Toronto
2003-2004	Robert and Jean Hadgraft Graduate Fellowship in Chemistry (\$4,000)	U. of Toronto
2000-2002	NSERC PGSB (\$38,200 total). <i>[The Canadian equivalent of the National Science foundation.]</i>	Canada
2001-2002	Edwin Walter Warren Graduate Student Award (\$4,000)	U. of Toronto
1998-2000	NSERC PGSA (\$34,200 total; foreign tenure granted but declined) <i>[The Canadian equivalent of the National Science foundation.]</i>	Canada
1998-1999	Eminent Scholar Award (\$3,500)	U. of Toronto
1998-2000	Alberta Heritage Foundation for Medical Research (\$35,000; declined)	Alberta
1998-1999	Ontario Graduate Scholarship (OGS, \$11,959; declined)	Ontario
1997-1998	Governor General's Silver Medal <i>[The silver Governor General's Academic Medal is awarded for top performance at the undergraduate level at a Canadian educational institution.]</i>	Brock University
1994-1998	Sears Canada Scholarship (\$10,000 total)	Canada
1994-1998	Donohue (QUNO) Scholarship (\$3,000 total)	Brock University
1994-1998	Canada Scholarship (\$10,000 total) <i>[a 4-year nationally competitive fellowship based on high school academic performance]</i>	Canada
1994-1998	Dean's Honours List	Brock University
1997-1998	Scholler Foundation Scholarship (\$1,700)	Brock University
1997-1998	John W. Bean and Kathryn Bean Becker Scholarship (\$1,350)	Brock University
1997-1998	The Chemical Institute of Canada Prize (silver medal)	Brock University
1997-1998	SCI Student Merit Award Winner (silver medal)	Brock University
1997-1998	Martin S. Gibson Scholarship (\$150)	Brock University
1996-1997	M.J. (Mel) Farquharson Scholarship (\$1,025)	Brock University
1994-1995	C. Boyd Slemon Scholarship (\$1,650)	Brock University
1994-1995	3M Scholarship in Chemistry (\$500)	Brock University

Named Fellowships

1999-2000	James C. Cumming Fellow , Trinity College, Canada (\$6,800). [<i>A competitive, application-based fellowship to serve as a Junior Fellow of Trinity College at the University of Toronto. The expectation was to contribute positively to the life of the college.</i>]	U. of Toronto
2000-2002	Walter C. Sumner Memorial Fellowship (\$9,000 total). [<i>A competitive, nomination and application-based fellowship at select Canadian universities for studies in chemistry, physics, electronics, and electrical engineering.</i>]	Canada

Publications

Notable Journal Impact Factors (2018):

- ACS Nano 13.903
- ACS Materials and Interfaces 8.456
- Journal of Physical Chemistry Letters 7.329
- Nature Communications 11.880
- Nature 43.070
- Science 41.04
- PNAS 9.412

Author order: Unless otherwise noted, Jason R. Dwyer as first or last author for work post-2009 denotes conception, development, and analysis of the reported research, and writing of at least the final version of the manuscript. My name is first for my papers published at URI when I performed a significant portion of the work independently of my collaborators.

Google Scholar: <https://scholar.google.com/citations?user=cFK7JhMAAAAJ>
3265 citations; h-index 19; i10-index 28.

^{UG}Denotes undergraduate author

1. **ACS Nano (2022)**. *Photoswitchable Binary Nanopore Conductance and Selective Electronic Detection of Single Biomolecules under Wavelength and Voltage Polarity Control*. James T. Hagan, Alejandra Gonzalez, Yuran Shi, Grace G. D. Han, and Jason R. Dwyer. <https://doi.org/10.1021/acsnano.1c10039>
2. **(CORRESPONDENCE) Nature (2021) 595, 30**. *Notice who the science system honours, and how*. Jason R. Dwyer. <https://doi.org/10.1038/d41586-021-01785-3>
3. **PNAS (2021) 118, e2022806118**. *Synthetic heparan sulfate standards and machine learning facilitate the development of solid-state nanopore analysis*. Ke Xia, James T Hagan, Li Fu, Brian S Sheetz, Somdatta Bhattacharya, Fuming Zhang, Jason R Dwyer, and Robert J. Linhardt. <https://doi.org/10.1073/pnas.2022806118>
4. **Review of Scientific Instruments (2021) 92, 043102**. *Targeting Improved Reproducibility in Surface-Enhanced Raman Spectroscopy with Planar Substrates Using 3D Printed Alignment Holders*. Buddini Iroshika Karawdeniya, Robert B. Chevalier, Y.M. Nuwan D.Y. Bandara, and Jason R. Dwyer. <https://doi.org/10.1063/5.0039946>
5. **Journal of Raman Spectroscopy (2021) 52, 608-615**. *Optimizing non-contact oxygen-plasma treatment to improve the performance of a top-down nanofabricated SERS substrate with structurally responsive, high-aspect-ratio nanopillar array*. Robert B. Chevalier and Jason R. Dwyer. <http://dx.doi.org/10.1002/jrs.6050>
6. **Analytical and Bioanalytical Chemistry (2020) 412, 6639-6654**. *Chemically Tailoring Nanopores for Single-Molecule Sensing and Glycomics*. James T. Hagan, Brian S. Sheetz, Y.M. Nuwan D.Y. Bandara, Buddini I. Karawdeniya, Melissa A. Morris,^{UG} Robert B. Chevalier, and Jason R. Dwyer. <https://doi.org/10.1007/s00216-020-02717-2>
7. **Nanotechnology (2020) 31 335707**. *Beyond Nanopore Sizing: Improving Solid-State Single-Molecule Sensing Performance, Lifetime, and Analyte Scope for Omics by Targeting Surface Chemistry During Fabrication*. Y. M. Nuwan D. Y. Bandara, Jugal Saharia, Buddini I. Karawdeniya, James T Hagan, Jason R. Dwyer, and Min Jun Kim. <https://doi.org/10.1088/1361-6528/ab8f4d>
8. **ACS Applied Nano Mater. (2020) 3, 2969-2977**. *Rapid, General-Purpose Patterning of Silicon Nitride Thin Films Under Ambient Conditions for Applications Including Fluid Channel and SERS*

- Substrate Formation*. Brian Sperry Sheetz, Y.M. Nuwan D.Y. Bandara, Benjamin Rickson,^{UG} Michael Auten,^{UG} and Jason R. Dwyer. <https://doi.org/10.1021/acsanm.0c00248>
9. **ACS Nano (2019) 13, 8155-8168.** *Geometry-Based Self-Assembly of Histone-DNA Nanostructures at Single-Nucleotide Resolution*. Maged F. Serag, Aimaiti Aikeremu, Ryoko Tsukamoto, Hubert Piwonski, Maram Abadi, Noritada Kaji, Jason R Dwyer, Yoshinobu Baba, and Satoshi Habuchi. <https://doi.org/10.1021/acsnano.9b03259>
 10. **Applied Spectroscopy (2019) 73, 1370-1379.** *An Open-Source, Iterative Dual-Tree Wavelet Background Subtraction Method Extended from Automated Diffraction Pattern Analysis to Optical Spectroscopy*. Robert B. Chevalier and Jason R. Dwyer. <https://doi.org/10.1177/0003702819871330>
 11. **ACS Materials and Interfaces (2019) 11, 30411-30420.** *Chemically Functionalizing Controlled Dielectric Breakdown Silicon Nitride Nanopores by Direct Photohydrosilylation*. Y. M. Nuwan D. Y. Bandara, Buddini I. Karawdeniya, James T. Hagan, Robert B. Chevalier, and Jason R. Dwyer. <https://pubs.acs.org/doi/10.1021/acsami.9b08004>
 12. **(INVITED) Journal of Analysis and Testing (2019) 3, 61-79.** *Challenging Nanopores with Analyte Scope and Environment*. Buddini I. Karawdeniya, Y.M. Nuwan D.Y. Bandara, Jonathan W. Nichols, Robert B. Chevalier, James T. Hagan, and Jason R. Dwyer. <https://doi.org/10.1007/s41664-019-00092-1>
 13. **ACS Omega (2019) 4, 226-230.** *A Push-Button Method to Create Nanopores Using a Tesla Coil Lighter*. Y. M. Nuwan D. Y. Bandara, Buddini I. Karawdeniya,^{1,†} and Jason R. Dwyer. <https://pubs.acs.org/doi/10.1021/acsomega.8b02660>
 14. (preprint) **ChemRxiv (2018).** *"Lights, Camera, Questions and Answers!"*: Talking About Science on Camera. Buddini Karawdeniya Y.M. Nuwan D.Y. Bandara, Julie Whelan, John Yacano,^{UG} Marissa DeOliveira,^{UG} Dana Neugent, Regina Bell, Jason R. Dwyer. <https://doi.org/10.26434/chemrxiv.7496294.v1>
 15. **Nature Communications (2018) 9, 3278.** *Surveying silicon nitride nanopores for glycomics and heparin quality assurance*. Buddini Iroshika Karawdeniya, Y.M. Nuwan D.Y. Bandara, Jonathan W. Nichols, Robert B. Chevalier, and Jason R. Dwyer. <https://doi.org/10.1038/s41467-018-05751-y>
 16. **ACS Appl. Nano. Mater. (2018) 1, 960-968.** A General Strategy to Make an On-Demand Library of Structurally and Functionally Diverse SERS Substrates. Buddini Iroshika Karawdeniya, Y.M. Nuwan D.Y. Bandara, Julie C. Whelan, and Jason R. Dwyer. <https://pubs.acs.org/doi/10.1021/acsanm.7b00385>
 17. **(INVITED) ELECTROPHORESIS (2018) 39, 626-634.** *Conductance-Based Profiling of Nanopores: Accommodating Fabrication Irregularities*. Y.M. Nuwan D.Y. Bandara, Jonathan W. Nichols, Buddini Iroshika Karawdeniya, and Jason R. Dwyer. <https://onlinelibrary.wiley.com/doi/full/10.1002/elps.201700299>.
 18. **Physical Chemistry Chemical Physics (2017) 19, 27074-27080.** *A Comparison of SERS and MEF of Rhodamine 6G on A Gold Substrate*. Elizabeth Kohr, Buddini I. Karawdeniya, Jason R. Dwyer, Anju Gupta, and William B Euler. <https://pubs.rsc.org/en/content/articlehtml/2017/CP/C7CP05569B>. [My group provided the means to prepare the part of the experiment responsible for optical enhancement, and we contributed to the writing of the paper.]
 19. **(INVITED, Focal Point Review) Applied Spectroscopy (2017) 71, 2051-2075.** Jason R. Dwyer and Maher Harb. *Through a window, brightly: A review of selected nanofabricated thin film platforms for spectroscopy, imaging, and detection*. DOI: <https://doi.org/10.1177/0003702817715496>
 20. **ACS Applied Materials & Interfaces (2016) 8, 34964-34969.** Y.M. Nuwan D.Y. Bandara, Buddini Iroshika Karawdeniya, Julie C. Whelan, Lucas D.S. Ginsberg,^{UG} and Jason R. Dwyer. *Solution-Based Photo-Patterned Gold Film Formation on Silicon Nitride*. <https://pubs.acs.org/doi/10.1021/acsami.6b12720>
 21. **(INVITED) Nanofluidics 2nd edition: Nanoscience and Nanotechnology Series (2016) 196-236.** Jason R. Dwyer*, Y.M. Nuwan D.Y. Bandara, Julie C. Whelan, Buddini Iroshika Karawdeniya, and Jonathon W. Nichols. *Silicon nitride membranes for nanofluidic device fabrication*. (Eds. Joshua Edel & Min Jun Kim, Royal Society for Chemistry). <http://dx.doi.org/10.1039/9781849735230>.
 22. **ACS Applied Materials & Interfaces (2016), 8, 30583-30589.** *Real-time Profiling of Solid-State Nanopores During Solution-Phase Nanofabrication*, Y.M. Nuwan D.Y. Bandara, Buddini Iroshika Karawdeniya, and Jason R. Dwyer. <https://pubs.acs.org/doi/10.1021/acsami.6b10045>
 23. **ACS Applied Materials & Interfaces (2014) 6, 10952-10957.** Julie C. Whelan, Buddini Iroshika Karawdeniya[‡], Y.M. Nuwan D.Y. Bandara[‡], Brian D. Velleco, Caitlin M. Masterson^{UG} and Jason R.

- Dwyer. *Electroless Plating of Thin Gold Films Directly onto Silicon Nitride Thin Films and into Micropores*. ‡equal contributions. <https://pubs.acs.org/doi/10.1021/am501971n>.
24. **(INVITED, cover) ACS Applied Materials & Interfaces Forum: New Frontiers and Challenges in Biomaterials (2013) 5, 9330-9337.** C.M. Frament,^{UG} N. Bandara and J.R. Dwyer, *Nanopore surface coating delivers nanopore size and shape through conductance-based sizing*. <https://pubs.acs.org/doi/10.1021/am4026455>.
 25. **(INVITED FOR ACS LIVESLIDES) J. Phys. Chem. Lett. (2013) 4, 2339.** C. Mueller, M. Harb, J.R. Dwyer and R.J.D. Miller. *Nanofluidic cells with controlled path length and liquid flow for rapid, high-resolution in situ Imaging with Electrons*. <https://pubs.acs.org/doi/10.1021/jz401067k>.
 26. **Review of Scientific Instruments (2013) 84, 036101-036101-2.** Jason R. Dwyer*, Łukasz Szycc, Erik T.J. Nibbering and Thomas Elsaesser. *Note: An environmental cell for transient spectroscopy on solid samples in controlled atmospheres*. *Corresponding author. <https://aip.scitation.org/doi/10.1063/1.4794092>
 27. **MRS Online Proceedings Library (2013) 1544.** C. Mueller, M. Harb, J.R. Dwyer and R.J.D. Miller. *Nanofluidic Cells with Controlled Path Length and Liquid Flow For Rapid, High-Resolution In Situ Electron Microscopy*. <http://dx.doi.org/10.1557/opl.2013.780>
 28. **Journal of Physical Chemistry C (2012) 116, 23315-23321.** C.M. Frament^{UG} and J.R. Dwyer. *Conductance-Based Determination of Solid-State Nanopore Size and Shape: An Exploration of Performance Limits*. <https://pubs.acs.org/doi/10.1021/jp305381j>.
 29. **Journal of Physical Chemistry A (2011) 115, 13149-13157.** R. Dawes*, J.R. Dwyer*, W. Qu and K.M. Gough (*equal contributions). *QTAIM Investigation of the Electronic Structure and Large Raman Scattering Intensity of Bicyclo-[1.1.1]-pentane*. <https://pubs.acs.org/doi/10.1021/jp205658z>
 30. **ACS Nano (2009) 3, 3009-3014.** V. Tabard-Cossa, M. Wiggin, D. Trivedi, N.N. Jetha, J.R. Dwyer and A. Marziali. *Single-Molecule Bonds Characterized by Solid-State Nanopore Force Spectroscopy*. <https://pubs.acs.org/doi/abs/10.1021/nn900713a>
 31. **(SPECIAL ISSUE) Chemical Physics (2009) 357, 36-44.** Ł. Szycc, J.R. Dwyer, E.T.J. Nibbering and T. Elsaesser. *Ultrafast dynamics of N-H and O-H stretching excitations in hydrated DNA oligomers*. <https://doi.org/10.1016/j.chemphys.2008.08.013>
 32. **Journal of Physical Chemistry B, (2008) 112, 11194-11197.** J.R. Dwyer, Ł. Szycc, E.T.J. Nibbering and T. Elsaesser. *Ultrafast vibrational dynamics of adenine-thymine base pairs in DNA oligomers*. <https://pubs.acs.org/doi/10.1021/jp8054119>
 33. **Springer Series in Chemical Physics, Ultrafast Phenomena XVI (2008) 92, 535-537.** J.R. Dwyer, Ł. Szycc, E.T.J. Nibbering and T. Elsaesser. *Ultrafast vibrational dynamics of adenine-thymine base pairs in hydrated DNA*.
 34. **(INVITED) K.M. Gough, R. Dawes, J.R. Dwyer and T.R. Welshman, QTAIM analysis of Raman scattering intensities: Insights into the relationship between molecular structure and electronic charge flow.** In *Recent Advances in the Quantum Theory of Atoms in Molecules* (Eds. R.J. Boyd and C. Matta., Wiley-VCH). March 2007, ISBN 978-3-527-30748-7. [Refereed book chapter]
 35. **(INVITED) Chemical Physics (2007) 341, 175-188.** T. Elsaesser, N. Huse, J. Dreyer, J.R. Dwyer, K. Heyne and E.T.J. Nibbering. *Ultrafast vibrational dynamics and anharmonic couplings of hydrogen-bonded dimers in solution*. <https://doi.org/10.1016/j.chemphys.2007.06.036>
 36. **(INVITED) Journal of Modern Optics (2007) 54, 923-952.** J.R. Dwyer, R.E. Jordan, C.T. Hebeisen, M. Harb, R. Ernstorfer, T. Dartigalongue and R.J.D. Miller. *Experimental basics for femtosecond electron diffraction studies*. (DOI: 10.1080/09500340601125020)
 37. **Journal of Modern Optics (2007) 54, 905-922.** J.R. Dwyer, R.E. Jordan, C.T. Hebeisen, M. Harb, R. Ernstorfer, T. Dartigalongue and R.J.D. Miller. *Femtosecond electron diffraction: An atomic perspective of condensed phase dynamics*. (DOI: 10.1080/09500340601095348)
 38. **Springer Series in Chemical Physics, Ultrafast Phenomena XV (2007) 88, 335-337.** A. Paarmann, D. Kraemer, M.L. Cowan, N. Huse, M. Harb, B.D. Bruner, J.R. Dwyer, E.T.J. Nibbering, T. Elsaesser and R.J.D. Miller. *2D-IR photon echo spectroscopy of liquid H₂O—Combination of novel nanofluidics and diffractive optics deciphers ultrafast structural dynamics*.
 39. **Chemical Physics Letters (2006) 432, 146-151.** J.R. Dwyer, J. Dreyer, E.T.J. Nibbering and T. Elsaesser. *Ultrafast dynamics of vibrational N-H stretching excitations in the 7-azaindole dimer*.
 40. **Philosophical Transactions of the Royal Society A (2006) 364, 741-778.** J.R. Dwyer, C.T. Hebeisen, R. Ernstorfer, M. Harb, V.B. Deyirmenjian, R.E. Jordan and R.J.D. Miller. *Femtosecond electron diffraction: "Making the molecular movie"*. (DOI: 10.1098/rsta.2005.1735)

41. **Nature (2005) 434, 199-202.** M.L. Cowan, B.D. Bruner, N. Huse, J.R. Dwyer, B. Chugh, E.T.J. Nibbering, T. Elsaesser and R.J.D. Miller. *Ultrafast memory loss and energy redistribution in the hydrogen bond network of liquid H₂O.* <https://www.nature.com/articles/nature03383> [9/27/2019: 561 citations]
42. **Springer Series in Chemical Physics, Ultrafast Phenomena XIV (2005) 79, 144-148.** J.R. Dwyer, R.E. Jordan, B.J. Siwick, C.T. Hebeisen and R.J.D. Miller. *Femtosecond electron diffraction: Towards making the "molecular movie"*.
43. **Chemical Physics (2004) 299, 285-305.** B.J. Siwick, J.R. Dwyer, R.E. Jordan and R.J.D. Miller. *Femtosecond electron diffraction studies of strongly driven structural phase transitions.*
44. **(JOURNAL COVER) Science (2003) 302, 1382-1385.** B.J. Siwick, J.R. Dwyer, R.E. Jordan and R.J.D. Miller. *An atomic level view of melting using femtosecond electron diffraction.* <https://science.sciencemag.org/content/302/5649/1382> [9/27/2019 630 citations]
45. **Springer Series in Chemical Physics, Ultrafast Phenomena XIII (2003), 71, 322-324.** B.J. Siwick, J.R. Dwyer, R.E. Jordan and R.J.D. Miller. *Ultrafast electron optics: Propagation dynamics and measurement of femtosecond electron packets.*
46. **Journal of Applied Physics (2003) 94, 807-808.** B.J. Siwick, J.R. Dwyer, R.E. Jordan and R.J.D. Miller. *Response to "Comment on 'Ultrafast electron optics: Propagation dynamics of femtosecond electron packets'"*.
47. **Journal of Applied Physics (2002) 92, 1643-1648.** B.J. Siwick, J.R. Dwyer, R.E. Jordan, and R.J.D. Miller. *Ultrafast electron optics: Propagation dynamics of femtosecond electron packets.*
48. **Canadian Journal of Chemistry (2000) 78, 1035-1043.** K.M. Gough, J.R. Dwyer and R. Dawes. *Ab initio analysis of C-H and C-C stretching intensities in Raman spectra of hydrocarbons.*
49. **Journal of Chemical Physics (1999) 111, 9971-9981.** M. Snajdr, J.R. Dwyer and S.M. Rothstein. *Histogram filtering: A technique to optimize wavefunctions for use in Monte Carlo simulations.*
50. **Journal of Physical Chemistry A (1998) 102, 2723-2731.** K.M. Gough and J.R. Dwyer. *Effect of structure and conformation on Raman trace scattering intensities in hydrocarbons.*
51. **Canadian Journal of Chemistry (1996) 74, 1139-1144.** K.M. Gough, M. Yacowar, R.H. Cleve and J.R. Dwyer. *Analysis of polarizability derivatives in H₂, HF, F₂, N₂ and CO with the theory of atoms in molecules.*

Intellectual Property

Issued Patents.

Patent # 10,351,958 (July 16, 2019). *Systems and methods for electroless plating of thin gold films directly onto silicon nitride and into pores in silicon nitride.* J.R. Dwyer, J.C. Whelan, Y.M. Nuwan D.Y. Bandara, and B.I. Karawdeniya.

Patent # 10,519,035 (Dec. 31, 2019). *Covalent chemical surface modification of surfaces with available silicon or nitrogen.* J.R. Dwyer, Y.M. Nuwan D.Y. Bandara, Buddini I. Karawdeniya, and J.C. Whelan.

Pending or Provisional Filings.

(provisional 5/8/2020) Surface Functionalization of Cellulose and Other Substrates.

United States Provisional Patent Application 62845040 (2019, University of Rhode Island): Cellulose-Based Substrate Modified with Naphthylethylenediamine

(provisional) High-Voltage Discharge to Roughen Wafer Surfaces for Applications Such as Surface-Enhanced Raman Spectroscopy (SERS)

Co-founder (2009): **Insight Nanofluidics**

Lectures and Presentations

Keynote

1. *Illuminating Single Molecule Science Without Light: The Role of Enquiry in Discovery and Application in Biomedical Diagnostics.* Showcasing Our Art and Research, St. Anselm College, Goffstown, NH, April 23, 2020.
2. *Shining a Light on Bay Chemistry and Technology Commercialization,* RI NSF EPSCoR Annual Research Symposium, University of Rhode Island, April 9, 2018.
3. *Nanopores: Portals to the Molecular World.* Chemical Biophysics Symposium, Toronto 2013, Canada, April 19-21, 2013. [International meeting; student-run, founded 2002]

Plenary

1. *Tailored Thin-Film Nanofluidics for Optical and Charged-Particle Single-Molecule Science*. Canadian Society for Chemistry Satellite Meeting on Ultrafast Spectroscopy and Imaging of Molecular Processes. University of Toronto, Toronto, ON, Canada. June 2, 2017. [International meeting]

Principal Speaker

1. *Synthetic nanopore force spectroscopy*. International Symposium: Advanced Science and Technology for Single Molecular Analysis of DNA and Related Molecules, Kyoto, Japan, January 24-26, 2011. [International meeting: to mark a major investment by the Japanese government in this technology, with invitations to internationally renowned speakers]

Award

1. *Chemical Approaches to Improve Nanopore Single-Molecule Sensing*. FACSS Innovation Award Finalist, SciX 2019, Palm Spring, California, October 17, 2019. [International meeting; *International conference established 1972, convened by a federation of 15 independent societies of analytical chemistry and spectroscopy, with strong academic and industrial representation.*]
2. *An atomic-level view of ultrafast structural dynamics using femtosecond electron diffraction*. Donald J. Le Roy Prize in Physical Chemistry Lecture, Department of Chemistry, University of Toronto, Toronto, Canada, April 27, 2005. [top prize for doctoral work in physical chemistry]

Invited Presentations

Notable Invitations

- 2021 Biophysical Society Meeting (Nanoscale Approaches to Biology Symposium)
- 2020 Telluride Workshop on Smart Polymers and Nuclear Pore Complexes
- 2020 Gordon Research Conference on Nanotechnology in Food & Agriculture
- Lilly Endowment Lecture, Dept. of Chemistry, University of Notre Dame, 2018
- American Chemical Society National Meetings
- FACSS SciX Annual Meetings – An international conference for academia and industry, organized by a federation of 15 independent societies engaged in chemical measurement.
- 231st Electrochemical Society Meeting
- 2015 Pittcon Conference & Expo
- Nanopores Conference 2012, Lanzarote, Spain
- 2011 Gordon Research Conference: Nucleosides, nucleotides & oligonucleotides
- Second International Conference on “Photo-Induced Phase Transitions, France, 2005
- Conference on Lasers and Electro-Optics Pacific Rim, Taipei, Taiwan, 2003

Selected University Invited Presentations

Rensselaer Polytechnic Institute, UMass Amherst, U Penn, U Kansas, Providence College, Brock University, U of Montreal, McGill University, Drexel, U of Manitoba, U Waterloo, Guelph U, Worcester Polytechnic Institute, Colorado State University, UMass Dartmouth, Boston University, Tufts University, U of New Hampshire, U of Wisconsin-Milwaukee, Rhode Island College, UMass Lowell, Simon Fraser University, Kavli Institute for Nanoscience (Delft University), U of Calgary, U of Western Ontario, Harvard, Christian Albrechts University.

1. *Robust Chemical Tailoring of Solid-State Nanopores for Glycomics (and -Omics, Generally)*. (International) Nanopore Weekly Meeting. Feb. 28, 2022. Online.
2. *Molecular-Level Tuning of Solid-State Nanopore Biomolecule Sensing: Optimizing DNA and Protein Analysis and Enabling Complex Carbohydrate Characterization*. 66th Biophysical Society Annual Meeting (Nanoscale Approaches to Biology Symposium), San Francisco, CA, Feb. 19-23, 2022.
3. *Electronic Single-Molecule Sensing for Glycomics and Genomics Using Chemically Tailored Nanopores*. Global GSK Talk (GSK Vaccines External R&D), Virtual, October 12, 2021.
4. *Tuning, Torturing, and Touching Up a SERS Substrate*. SciX2021, Providence, RI, USA, Sept. 26-Oct.1 2021.
5. *Rapid and direct single-molecule sensing, characterization, and sequence-level profiling of complex carbohydrates*. 5th New England Glycochemistry Meeting. Virtual meeting, July 16, 2021.
6. *Chemically Tuned Silicon Nitride Nanopores for Nucleic Acid Sequencing*. NIH NHGRI Advanced Genomic Technology Development Meeting. Virtual meeting, May 27-29, 2021.

7. *Customizing Solid-State Nanopores and Nanopore-Omics*. 2020 Telluride Workshop on Smart Polymers and Nuclear Pore Complexes, Telluride, CO, June 8-12, 2020 [Postponed to 2022 due to COVID-19]
8. *Nanostructured devices for sensing: From Optical Nitrate and Nitrite Detection to Electronic Single-Molecule Glycan Profiling*. 2020 Gordon Research Conference on Nanotechnology in Food & Agriculture, Southern New Hampshire University, Manchester, NH, June 21-26, 2020. [International meeting] [Postponed due to COVID-19]
9. *Illuminating Single Molecule Science Without Light: The Role of Enquiry in Discovery and Application in Biomedical Diagnostics*. Showcasing Our Art and Research Event, St. Anselm College, Manchester, NH, April 23, 2020. [Postponed due to COVID-19].
10. *Single-Molecule –Omics Using Nanopores*. School of Molecular Sciences, Arizona State University, Tempe, AZ, March 31, 2020. [Cancelled due to COVID-19]
11. *Single-Molecule –Omics Using Nanopores*. Department of Chemistry and Chemical Biology, IUPUI, Indianapolis, IN, March 25, 2020. [Postponed due to COVID-19]
12. *Delving Into the Nanoscale World With Thin-Film Nanofluidic Devices*. Department of Chemistry, College of the Holy Cross, Worcester, MA, April 3, 2020. [Postponed due to COVID-19]
13. *Delving Into the Nanoscale World with Thin-Film Nanofluidic Devices*. Department of Chemistry, Rensselaer Polytechnic Institute, Troy, NY, Oct. 22, 2019.
14. *Chemical Approaches to Improve Nanopore Single-Molecule Sensing*. FACSS Innovation Award Finalist, SciX 2019, Palm Spring, California, October 17, 2019.
15. *Expanding Nanopore Scope to Polysaccharide Characterization and Augmenting Performance by Surface Chemical Modifications*. Polymer Science and Engineering Department, University of Massachusetts, Amherst, MA, October 1, 2019.
16. *Nanopore Single Molecule Sensing and Platform Development*. Department of Chemistry, University of Rhode Island, Kingston, RI, April 1, 2019.
17. *Delving Into the Nanoscale World with Thin-Film Nanofluidic Devices*. Department of Physics (Condensed Matter), University of Pennsylvania, Philadelphia PA, March 27, 2019.
18. Tech Talk – Sensor Development: Coastal Ecology Assessment, Innovation, and Modeling. The Southeastern New England Defense Industry Alliance, Charlestown, RI, USA, Feb. 14, 2019.
19. **(Lilly Endowment Lecture)** *Nanopore Single Molecule Sensing and Platform Development*. Department of Chemistry and Biochemistry, University of Notre Dame, South Bend, IN, Oct. 8, 2018.
20. *Thin-Film Nanofluidic Devices for Single-Molecule Science: Electronic, Optical, and Force Sensor Platforms*. Department of Chemistry, University of Kansas, Lawrence, KS, USA, Sept. 28, 2018.
21. *Detecting Single Molecules and Their Interactions Using Nanopores*. Department of Chemistry and Biochemistry, Providence College, Providence, USA, Sept. 12, 2018.
22. *Single-molecule and interaction-based biosensing using nanopores: Oligo and polysaccharide analysis*, American Chemical Society, 256th National Meeting, Boston, USA, August 19-23, 2018. [International meeting]
23. *Chemically tuned thin-film and nanofluidic sensors for single-molecule and particle sensing*, American Chemical Society 255th National Meeting, New Orleans, USA, March 18-22, 2018. [International meeting]
24. *Single Molecule Biopolymer Analysis Using Interface-Tailored Nanopore Sensing*, The U.S. Army Natick Soldier Research, Development & Engineering Center, Natick, MA, Dec. 7, 2017.
25. *Thin-film nanofluidics for single-particle analysis*, American Chemical Society 254th National Meeting, Washington, D.C., August 20-24, 2017. [International meeting]
26. *Thin-Film Nanofluidic Devices for Single-Molecule Science: Electronic, Optical, and Force Sensor Platforms*. Department of Chemistry, Brock University, St. Catharines, ON, Canada. June 5, 2017.
27. *Thin-Film Nanofluidic Devices for Single-Molecule Science: Electronic, Optical, and Force Sensor Platforms*. 231st ECS Meeting (The Electrochemical Society), New Orleans, LA. May 28-June 1, 2017.
28. *Thin-Film Nanofluidic Devices for Single-Molecule Science: Electronic, Optical, and Force Sensor Platforms*. Department of Chemistry, University of Montreal, Montreal, QC, Canada. April 7, 2017. [International meeting]
29. *Thin-Film Nanofluidic Devices for Single-Molecule Science: Electronic, Optical, and Force Sensor Platforms*. Department of Chemistry, McGill University, Montreal, QC, Canada. April 6, 2017.

30. *Pores, Particles, Fibres, and Films: Tailored Nanostructures for Charged-Particle Single-Molecule Sensing and Spectroscopy*. Department of Physics, Drexel University, Philadelphia, PA. January 26, 2017.
31. *Pores, Particles, Fibres, and Films: Tailored Nanostructures for Charged-Particle Single-Molecule Sensing and Spectroscopy*. Department of Chemistry, University of Manitoba. July 28, 2016.
32. *Pores, Particles, Fibres, and Films: Tailored Nanostructures for Charged-Particle Single-Molecule Sensing and Spectroscopy*. Department of Chemistry, University of Waterloo, Waterloo, Canada, June 15, 2016.
33. *Pores, Particles, Fibres, and Films: Chemically tailoring nanostructures for Exploring the Molecular World*. Department of Chemistry, Guelph University, Guelph, Canada, June 13, 2016.
34. *Nanochannel sensing devices enhanced with tailored thin films: applications to single-molecule bioanalysis and materials characterization*. Third Arab American Frontiers, of Science, Engineering, and King Abdullah University of Science and Technology (KAUST) Medicine Symposium, Thuwal, Saudi Arabia (declined attendance). [International meeting]
35. *Clowns in a car: Nanofluidics for single-molecule science*. Chemistry and Biochemistry Department, Oberlin College, Oberlin, OH, March 30, 2016.
36. *Molecular-Level Design of Nanoscale Tools for Enhanced Single-Molecule Sensing*. FACSS SciX 2015, Providence, RI, [Julie C. Whelan](#), Y.M. Nuwan D.Y. Bandara, Buddini Iroshika Karawdeniya, and Jason R. Dwyer.
37. *Tailored silicon nitride thin-films for optical and all-electronic chemical sensing*. 250th ACS National Meeting, August 16-20, 2015, Boston, MA, USA. [International meeting]
38. *Thin-membrane nanochannels for nanopore single-molecule sensing and transmission electron microscopy of liquid samples*. Pittcon Conference & Expo, March 8-12, 2015, New Orleans, LA, USA [International meeting; the premier conference for chemical instrumentation]
39. *Nanofluidics for single particle imaging and sensing*. Chemistry seminar, Worcester Polytechnic Institute, October 1, 2014.
40. *Liquid electron microscopy and nanopore sensing: Two portals to the molecular world*. Physikalisch-Technische Bundesanstalt (National Metrology Institute of Germany), Berlin, Germany, July 21, 2014.
41. *Nanopore sensors and liquid TEM cells*. NIST Research Group Seminar, Boulder, CO, March 13, 2014 (invited, could not deliver the talk).
42. *Liquid electron microscopy and nanopore sensing: Two portals to the molecular world*. Analytical Chemistry Seminar, Department of Chemistry, Colorado State University, Ft. Collins, CO, March 12, 2014.
43. *Nanoscience and Nanotechnology*. Osher Lifetime Learning Institute, Kingston, RI, November 6, 2013.
44. *Probing Nanoscale Structure and Interaction in Highly Constrained Environments and in Free Solution: Nanopore Force Spectroscopy and Liquid Electron Microscopy*, Chemistry Seminar, Department of Chemistry, University of Massachusetts, Dartmouth, Oct. 23, 2013.
45. *Probing Nanoscale Structure and Interaction in Highly Constrained Environments and in Free Solution: Nanopore Force Spectroscopy and Liquid Electron Microscopy*, Physical Chemistry Seminar, Department of Chemistry, Boston University, Boston, MA, Oct. 9, 2013.
46. *Nanofabricated sensors and electron microscopy on submerged samples*. University of Rhode Island School of Oceanography Marine and Atmospheric Chemistry Seminar, Narragansett, RI, Oct. 4, 2013.
47. *Nanofluidic cell for high-resolution, in-liquid transmission electron microscopy*. Osher Lifetime Learning Institute, Kingston, RI, May 1, 2013.
48. *Through the ultrathin window and into the nanoscale hole: I. Transmission electron microscopy (TEM) of liquid-phase samples and II. Nanopore force spectroscopy of molecular interactions*. Department of Chemistry, Tufts University, Medford, MA, January 29, 2013.
49. *Nanopores and Nanofluidics: Molecular imaging and manipulation*. Nabsys, Inc., Providence RI, January 17, 2013.
50. *Through the ultrathin window and into the nanoscale hole: I. Transmission electron microscopy (TEM) of liquid-phase samples and II. Nanopore force spectroscopy of molecular interactions*. Department of Chemistry, University of New Hampshire, Durham, NH, April 4, 2013
51. *Single nucleotide resolution using nanopore force spectroscopy*. GTC 2nd Next Generation Sequencing Conference, Boston, USA, May 30-31, 2012. [International meeting]

52. *Nanopores: Portals to the Molecular World*. Department of Physics, University of Wisconsin-Milwaukee, May 4, 2012.
53. *Solid-state nanopore force spectroscopy and in situ nanopore sizing—performance limits*. Nanopores Conference 2012, Lanzarote, Spain, Feb. 6-10, 2012. [International meeting]
54. *Building and using the molecular-scale laboratory*. Rhode Island College, Providence, RI, January 27, 2012.
55. *Nanotechnology: Forging a better future one molecule at a time*. URI Honors Colloquium, University of Rhode Island, October 18, 2011.
56. *Biosensing with Nanopore Force Spectroscopy*. Analytical Chemistry Chemistry Seminar, Department of Chemistry, University of Massachusetts, Lowell. Sept. 21, 2011.
57. *Nanopore sequencing and the demonstration of single base pair resolution*. Gordon Research Conference: Nucleosides, nucleotides & oligonucleotides, Newport, RI, July 3-8, 2011. [International meeting]
58. *Nanopores: Portals to the Molecular World*. Amgen Seminar Series in Chemical Engineering, University of Rhode Island, Kingston, RI. February 17, 2011.
59. *Nanotechnology*. Oceanography 2030. W. Alton Jones Campus, URI. West Greenwich, RI. January 12-13, 2011.
60. *Nanopores: Portals to the Molecular World*. Department of Chemistry Seminar Series, University of Rhode Island, October 5, 2009.
61. *Synthetic Nanopore Force Spectroscopy*. NHGRI 2009 Sequencing Technology Development Grantee Meeting (Closed Meeting), La Jolla, California, March 29-31, 2009.
62. *Hydrogen bonding in DNA: Ultrafast vibrational dynamics of base pairs and waters of hydration*. Biophysics Seminar, Simon Fraser University, Burnaby, British Columbia, January 12, 2009.
63. *Strongly-driven melting of metals viewed in real-time and real space using femtosecond electron diffraction*. Bereich Seminar A, Max-Born-Institute, Berlin, Germany, October 25, 2007.
64. *Molecular movie-making: Capturing ultrafast structural dynamics with femtosecond infrared spectroscopy and femtosecond electron diffraction*. Atomic, Molecular and Optical Physics Seminar, Department of Physics, University of British Columbia, Vancouver, Canada, Oct. 2, 2007.
65. *Molecular movie-making: Capturing ultrafast structural dynamics with femtosecond infrared spectroscopy and femtosecond electron diffraction*. Sonderseminar des Centrums für Angewandte Photonik. Universität Konstanz, Konstanz, Germany, July 6, 2007.
66. *Molecular movie-making: Capturing ultrafast structural dynamics with femtosecond infrared spectroscopy and femtosecond electron diffraction*. Kavli Institute of Nanoscience, Delft University of Technology, Delft, The Netherlands, April 27, 2007.
67. *Molecular movie-making: Capturing ultrafast structural dynamics with femtosecond infrared spectroscopy and femtosecond electron diffraction*. Department of Chemistry, University of Calgary, Calgary, Canada, March 15, 2007.
68. *Ultrafast structural dynamics: From vibrational dynamics in DNA base pair analogues with femtosecond infrared spectroscopy to atomic-level movies of melting using femtosecond electron diffraction*. Departments of Chemistry and Physics, University of Western Ontario, London, Canada, March 7, 2007.
69. *Ultrafast structural dynamics: From vibrational dynamics in DNA base pair analogues with femtosecond infrared spectroscopy to atomic-level movies of melting using femtosecond electron diffraction*. Department of Chemistry, Harvard University, Boston, U.S.A., December 14, 2006.
70. *Structure and Function: Studies with Femtosecond Infrared Spectroscopy and Femtosecond Electron Diffraction*. Temps Group, Institute for Physical Chemistry, Christian Albrechts University, Kiel, Germany, November 6, 2006.
71. *Femtosecond electron diffraction: A direct probe of ultrafast structural dynamics*. Second International Conference on “Photo-Induced Phase Transitions: Cooperative, Non-linear and Functional Properties” (PIPT), University of Rennes, France, May 24-28, 2005. [International meeting]
72. *Femtosecond electron diffraction: A direct probe of strongly-driven melting dynamics in metals*. The Study of Matter at Extreme Conditions Conference (SMEC) 2005, Miami, Florida, April 17-21, 2005.
73. *Femtosecond electron diffraction: A tool to probe ultrafast structural dynamics*. Swiss Light Source, Villigen, Switzerland, January 17, 2005.
74. *Femtosecond electron diffraction: The quest for the molecular movie*. Department of Chemistry, Brock University, Canada, September 10, 2004.

75. *Making movies of molecules with femtosecond electron wavepackets*. Conference on Lasers and Electro-Optics Pacific Rim, Taipei, Taiwan, December 15-19, 2003. [International meeting]

Contributed Presentations

1. *Electronic Single-Molecule Sensing for Glycomics and Genomics Using Chemically Tailored Nanopores*. SciX2021, Providence, RI, USA, Sept. 26-Oct.1 2021.
2. *Nanopore Sensors for -omics: Combing Through the Microbiome one Molecule at a Time*. Rhode Island Microbiome Symposium 2020, Kingston, RI, Jan. 16-17, 2020.
3. *Fabrication and Analysis Tools for Real-Time Environmental Monitoring by Surface-Enhanced Raman Spectroscopy (SERS)*. SciX 2019, Palm Spring, California, October 16, 2019.
4. *Chemically Tuned Nanopore Sensor Platforms for Single-Molecule Sensing*. FACSS SciX 2018, Atlanta, GA, USA, October 26-26, 2018. [International meeting]
5. *Nanopore Single-Molecule Sensors for Oligo- and Polysaccharide Analysis*. FACSS SciX 2018, Atlanta, GA, USA, October 26-26, 2018. [International meeting]
6. *Single Molecule Glycomics Using Interface-Tailored Nanopore Sensing*, Gordon Conference on Bioanalytical Sensors, Newport, RI, June 24-29, 2018. (poster)
 - a. [+3 graduate student presentations]. *Characterization and Application of Surface Modified Solid-State Nanopores*, Robert B. Chevalier, Buddini I. Karawdeniya, Y. M. Nuwan D. Y. Bandara, James Hagan, Jonathan W. Nichols, and Jason R. Dwyer
 - b. *Sweets to Nutrients, We Keep Track of All: Single Molecule Sensing and Molecular Fingerprinting*. Buddini Iroshika Karawdeniya, Y.M. Nuwan D.Y. Bandara, Jonathan W. Nichols, Robert B. Chevalier, Julie C. Whelan and Jason R. Dwyer
 - c. *Customization of Solid State Nanopores for Single Molecule Detection*. James Hagan, Y.M. Nuwan D.Y. Bandara, Jason R. Dwyer.
7. *Low-Overhead Thin-Film Approaches and Platforms for Spectroscopic Fingerprinting and Electronic Single-Molecule Sensing*. 231st ECS Meeting (The Electrochemical Society), New Orleans, LA. May 28-June 1, 2017. [International meeting]
8. *Pores, Particles, Fibres, and Films: Tailored Nanostructures for Charged-Particle Single-Molecule Sensing and Spectroscopy*. Gordon Research Conference on Bioanalytical Sensors (From Atoms to Organisms: Bioanalytical Sensors Elucidating Physicochemical Properties of Multiscale Systems). Newport, RI, June 26-July 1, 2016. [poster presentation] [International meeting]
 - a. [+3 graduate student presentations at Gordon Research Symposium June 25-26 & GRC] *Surface tuning of silicon nitride for optical and conductance based nanopore sensing*, Y. M. Nuwan D. Y. Bandara, Buddini I. Karawdeniya, Julie C. Whelan and Jason R. Dwyer.
 - b. *Biopolymer profiling by nanopore sensors*, Buddini I. Karawdeniya, Y.M. Nuwan D.Y. Bandara, Julie C. Whelan and Jason R. Dwyer.
 - c. *Electroless plating to tune the physiochemical properties of nanostructured silicon nitride for use in device fabrication*, Julie C. Whelan, Y.M. Nuwan D.Y. Bandara, Buddini I. Karawdeniya, Jason R. Dwyer.
9. FACSS SciX 2015, Providence, RI, 2 graduate student presentations
 - a. *Photopatterned Electroless Gold Deposition: Optimizing Film Patterning and Nanoscale Structure for Applications*. Y.M. Nuwan D.Y. Bandara, Buddini I. Karawdeniya, Julie C. Whelan, and Jason R. Dwyer.
 - b. [poster award winner] *Electroless gold plating as an adaptable tool to fabricate custom surface enhanced Raman spectroscopic (SERS) substrates*. Buddini Iroshika Karawdeniya, Y.M. Nuwan D.Y. Bandara, Caitlin M. Masterson, Julie C. Whelan, Brian D. Velleco, and Jason R. Dwyer.
10. 250th ACS National Meeting, August 16-20, 2015, Boston, MA, USA, 3 graduate student presentations. [International meeting]
 - a. *Electroless plating as a flexible tool for the creation of custom surface enhanced Raman spectroscopic (SERS) substrates*. Buddini I. Karawdeniya, Y. M. Nuwan D. Y. Bandara, Caitlin M. Masterson, Julie C. Whelan, and Jason R. Dwyer
 - b. *Electroless Plating of Thin Gold Films Directly onto Silicon Nitride Thin Films and into Micro- and Nanopores*. Julie C. Whelan, Y. M. Nuwan D. Y. Bandara, Buddini I. Karawdeniya, and Jason R. Dwyer.

- c. *Photoinitiated covalent surface functionalization for enhanced control over electroless deposition on silicon nitride.* Y. M. Nuwan D. Y. Bandara, Buddini I. Karawdeniya, Brian Velleco, Julie Whelan, and Jason R. Dwyer.
11. Pittcon Conference & Expo, March 8-12, 2015, New Orleans, LA, USA , 1 undergraduate poster presentation [International meeting]
 - a. *Photochemical and Thermal Control over Electrolessly Gold-plated Film Structure on Thin Silicon Nitride to Target Sensing Applications.* Caitlin M. Masterson, Y.M. Nuwan D.Y. Bandara, Brian D. Velleco, Julie C. Whelan, Buddini Iroshika Karawdeniya, Jason R. Dwyer.
12. *Probing Nanoscale Structure and Interaction in Highly Constrained Environments and in Free Solution: Nanopore Force Spectroscopy and Liquid Electron Microscopy.* Gordon Research Conference on Single Molecule Approaches to Biology (Understanding Life at a Higher Resolution). Lucca, Italy, July 13-18, 2014. [poster presentation] [International meeting]
13. *Ultrathin membrane devices for diverse, high-performance nanoscale detection applications.* Gordon Research Conference on Bioanalytical Sensors (Twenty First Century Technologies for Probing Biological Systems). Newport, RI, June 22-27, 2014. [poster presentation] [International meeting]
 - a. [+4 graduate student poster presentations] *Modern Alchemy: Electroless Plating of Gold onto Silicon Nitride.* Julie C. Whelan, Buddini Iroshika Karawdeniya, Y.M. Nuwan D.Y. Bandara, Brian D. Velleco, Sarah Golden, Caitlin Masterson, Jason R. Dwyer.
 - b. *Photopatterned Electroless Gold Deposition on Silicon Nitride: Enhanced Control Over a Powerful Sensing Platform Interface.* Nuwan Bandara, Buddini Karawdeniya, Julie Whelan, Brian Velleco, Jason R. Dwyer.
 - c. *1. Harnessing biology for single-molecule sensing; 2. Electroless Gold-Plated Silicon Nitride Chips: An Adventitious Substrate for SERS.* Buddini I. Karawdeniya, Julie Whelan, Y.M. Nuwan D.Y. Bandara, Brian Velleco, Caitlin Masterson, Caitlin R. Ross, Jason R. Dwyer.
 - d. (student award winner) *SERS Signal Degradation of Thin Gold Films on Silicon Nitride.* Brian Velleco, Buddini Karawdeniya, Nuwan Bandara, Julie Whelan, Caitlin Masterson, Sarah Golden and Jason R. Dwyer.
14. *Using conductance to characterize surface-coated, single-molecule nanopore sensor interfaces.* 245th ACS National Meeting, New Orleans, LA, April 7-11, 2013. [International meeting]
15. *Nanofluidic cell for high-resolution, in-liquid transmission electron microscopy.* 245th ACS National Meeting, New Orleans, LA, April 7-11, 2013. [International meeting]
16. *Characterizing single-molecule nanopore sensors: Performance limits of conductance-based methods.* 244th ACS National Meeting, Philadelphia, PA, August 19-23, 2012. [International meeting]
17. *Exploring the dramatic dependence of ssDNA:histone assembly on oligomer length.* 244th ACS National Meeting, Philadelphia, PA, August 19-23, 2012. [International meeting]
18. *Nanopore force spectroscopy for bioanalysis and biophysics.* Gordon Conference: Bioanalytical Sensors. Newport, RI, June 17-22, 2012. [International meeting]
19. *Solid-State Nanopore Force Spectroscopy.* NHGRI Advanced Sequencing Technology Grantee Meeting and NHGRI Advanced Sequencing Technology Development Meeting. San Diego, CA, April 4-6 and 6-7, 2011.
20. *Synthetic nanopore force spectroscopy for clinical genotyping.* ACS Fall 2010 National Meeting, Boston, MA, August 22-26, 2010. [International meeting]
21. *Nanopore Force Spectroscopy for Genotyping Applications.* Department of Physics Seminar Series, University of Rhode Island, November 20, 2009.
22. *Artificial nanopores for biophysical investigations and bioanalytical applications.* Frontiers in Biophysics 2009, Simon Fraser University, Burnaby, British Columbia, January 23, 2009.
23. *Nanopore force spectroscopy.* BioFest 2009, UBC, January 16, 2009.
24. *Molecular movie-making: Capturing ultrafast structural dynamics with femtosecond infrared spectroscopy and femtosecond electron diffraction.* Chen Group, Department of Chemistry, Cornell University, Ithaca, USA, September 28, 2007.
25. *Ultrafast N-H vibrational dynamics in the DNA model base pair 7-azaindole dimer.* Thirteenth International Conference on Time-Resolved Vibrational Spectroscopy. Freising, Germany, May 19-25, 2007. [International meeting]

26. *Ultrafast dynamics of vibrational N-H stretching excitations in the 7-azaindole dimer*. Deutschen Physikalischen Gesellschaft, Duesseldorf, Germany, March 19-23, 2007. [International meeting]
27. *Ultrafast melting in metals probed with femtosecond electron diffraction*. XX Congress of the International Union of Crystallography (IUCr2005), Florence, Italy, August 23-31, 2005 [J.R. Dwyer, R.E. Jordan, C.T. Hebeisen, M. Harb, R. Ernstorfer, and R.J.D. Miller, poster]. [International meeting]
28. *Femtosecond electron diffraction: A direct probe of ultrafast structural dynamics*. German-Canadian Workshop: Young Scientists in Photonics, Munich, Germany, June 10-12, 2005. [International meeting]
29. *Femtosecond electron diffraction: A tool to probe ultrafast structural dynamics*. Max Born Institute for Nonlinear Optics and Short Pulse Spectroscopy, Berlin, Germany, January 25, 2005.
30. *Ultrafast melting dynamics probed with femtosecond electron diffraction*. Institute of Physical Chemistry, University of Zurich, Zurich, Switzerland, January 21, 2005.
31. *Femtosecond electron diffraction: A tool to probe ultrafast structural dynamics*. Molecular Physics and Physical and Physical Chemistry Seminars, Laboratoire de Spectroscopie Ultrarapide, École Polytechnique Fédérale de Lausanne, Lausanne, Switzerland, January 18, 2005
32. *Femtosecond electron diffraction: The quest for the molecular movie*. Scherer Research Group, University of Chicago, Chicago, USA, January 7, 2005.
33. *Structural dynamics via femtosecond electron diffraction: The nature of ultrafast laser-induced melting*. 87th Canadian Chemistry Conference, London, Canada, May 29-June 1, 2004 [J.R. Dwyer, R.E. Jordan, C.T. Hebeisen, B.J. Siwick and R.J.D. Miller, poster].
34. *Femtosecond electron diffraction: Ultrafast melting dynamics revealed*. Canada-Germany Young Scientists in Photonics Workshop, Ottawa, Canada, September 2-3, 2003. [International meeting]
35. *Femtosecond electron diffraction: A direct probe of ultrafast structural dynamics*. The 39th IUPAC Congress and the 86th Conference of the Canadian Society for Chemistry, Ottawa, Canada, August 10-15, 2003. [International meeting]
36. *Ultrafast electron diffraction: Towards a structural probe of femtosecond molecular reaction dynamics*. University of Toronto Chemical Biophysics Symposium, Toronto, Canada, April 12-14, 2002 [J.R. Dwyer, B.J. Siwick, R.E. Jordan and R.J.D. Miller, poster].
37. *Ultrafast electron diffraction: A tool to probe reaction dynamics*. University of Toronto Physical Chemistry Seminar Series, Toronto, Canada, January 16, 2001.
38. *Ultrafast electron diffraction: Probing structural dynamics on the femtosecond timescale*. Photonics Research Ontario Student/Industry Retreat, Niagara-on-the-Lake, Canada, February 25-26, 2001 [B.J. Siwick, J.R. Dwyer, R.E. Jordan and R.J.D. Miller, poster].
39. *Ultrafast electron diffraction: Towards a direct probe of reaction dynamics*. 84th Conference of the Canadian Society for Chemistry, Montreal, Canada, May 26-30, 2001 [J.R. Dwyer, B.J. Siwick, R.E. Jordan and R.J.D. Miller, poster].
40. *Ultrafast electron diffraction: Reaction dynamics at the atomic level*. 16th Annual Symposium on Chemical Physics, Waterloo, Canada, November 3-5, 2000 [J.R. Dwyer, C. Belzile, B.J. Siwick, R.E. Jordan and R.J.D. Miller, poster].
41. Undergraduate poster presentations

Grant Funding

1. Champlin Foundation, 1/21-12/22, \$185,000, J. Menon (PI), J. Shen, G. Bothun, X. Chen, JRD, D. Roxbury. *Growing New Organs: Tissue Engineering Technologies for Hands-on Learning and Research*.
2. NIH R21 1R21HG011096-01, 4/20-3/2022, \$503,184 (\$349998 direct), Dwyer (PI). *Chemically Tuned Silicon Nitride Nanopores for Nucleic Acid Sequencing*.
3. Champlin Foundation, 2018, Dugan Hayes (PI), JRD, Leonard Kahn, Michael Tammaro, Daniel Roxbury, Tao Wei, \$175k. *Molecular Movies for the Selfie Generation: Nanosecond Laser Spectroscopy in Undergraduate STEM Education*.
4. URI ASU Innovation Hub Collaborative Research Seed Grant 2020, 7/1/2019-6/30/2020, Jason R. Dwyer (co-PI); Mark Hayes (ASU co-PI), \$50,000. *Sensitive and Selective All-Electronic Chemical and Biological Sensors for Smart Cities and Broadened Monitoring Capacities*.
5. NSF CHE-1808344, 8/1/2018-7/31/2022 Jason R. Dwyer (PI), \$349,075. *Gauging and Optimizing Solid-State Nanopore Sensing Performance for Polysaccharide Sensing and Glycomics*

6. Rhode Island Research Alliance (RI Science & Technology Advisory Council), Jason R. Dwyer (PI), \$80,000. *Engineering Reliable and Affordable Tools for Environmentally- and Economically-Driven Aquatic Monitoring*. January 2019-December 2019
7. NSF EPSCoR OIA-1655221, Geoff Bothun (Chem Eng, PI); Jason R. Dwyer - Thrust Lead overseeing ~\$300k/year spread amongst ~6 thrust investigators; ~\$50k direct to Dwyer/year, September 2017-August 2022, \$19M. *Rhode Island Consortium for Coastal Ecology Assessment, Innovation, and Modeling*. See <https://web.uri.edu/rinsfepscor/leadership/>
8. Champlin Foundation, 2017, Mindy Levine (PI), Jason R. Dwyer, Shahla Yekta, Michael McGregor, Cindy Graham Brittain, Sue Geldardt, Silvana Ngo, Arijit Bose, Thomas Boving, \$162,000, *Chemistry from the Back Row: Engaging Students Using a Suite of State-of-the Art Chemical Instruments for the Real-Time Visualization of Chemical Reactions and Phenomena*.
9. Rhode Island Research Alliance (RI Science & Technology Advisory Council), May 2017-May 2018, Jason R. Dwyer (PI), \$90,000. *Nanofluidic Profiling of Marine Biopolymers: Synthetic and Sensing Tools for Environmentally- and Economically-Driven Aquatic Monitoring*.
10. URI Council for Research Proposal Development Grant, July 2017-June 2018, Jason R. Dwyer (PI), \$14,250. *A Noninvasive Treatment for Resurrecting Biofouled Single-Molecule Nanofluidic Sensors*.
11. NSF CAREER AWARD CBET-1150085, May 2012-May 2018, Jason R. Dwyer (PI), \$400,000 (capped by program). *Enhancing molecular recognition biosensing with nanopore force measurements*.
12. Champlin Foundations, 2015, Xinyuan Chen (PI), Wei Lu, Samantha Meenach, Bingfang Yan, Geoffrey Bothun, Yi Zheng, Jason R. Dwyer, \$170,000. *Microfabrication Technology: An Urgent Need for Hands-On Learning at the University of Rhode Island*.
13. Champlin Foundations, Geoff Bothun (co-PI), Arijit Bose (co-PI), Jason R. Dwyer, Vinka Oyanedel-Craver, David Worthen, *An Advanced Hyperspectral Imaging System to Observe Nanoscale Materials and Processes across STEM*, \$154,000 (2014).
14. NIH R01 subcontract, *Nanopore array force spectroscopy chip for rapid clinical genotyping*, \$56,363 (direct costs).
15. Champlin Foundations, Jason R. Dwyer (PI), Mindy Levine, Geoff Bothun, Abraham Kovoov, *Advanced Instrumentation for Probing Structure and Physiological Function of Purified Target Molecules*, \$130,000 [awarded 2013/2014].
16. Senior Investigator NSF 1330406 *Collaborative Research: North East Water Resources Network*, (PI Art Gold, URI) \$2,000,000; \$200k to JRD. 08/01/2013-07/31/2017.
17. NSF REU Supplement for CBET 1150085, ~\$8,000 (2013)
18. URI Council for Research Proposal Development Grant, *Lowering the barriers to nanofabrication of nanopore single-molecule biomedical sensors*, July 2013-2014, \$15,000.
19. Rhode Island Nanotechnology Consortium, *A Nanofluidic Single-Molecule Characterization and Control Platform* (Dwyer, Keunhan Park & Radha Narayanan), 2012-2013, \$8000.
20. Champlin Foundations, Brenton DeBoef, Cindy Brittain, Jason R. Dwyer, Geoff Bothun, *A High Performance Liquid Chromatography—Mass Spectrometry Instrument*, \$142,566. [2010/2011]
21. Rhode Island Foundation, *Developing a low-cost, nanofabricated device for early, bedside cancer diagnosis*, \$15,000.
22. University of Rhode Island Council for Research, *Molecular analysis by nano-ruler: Using nanoparticle swelling to identify chemical species*, \$10,000.
23. Faculty start-up funds (URI)
 - o Rhode Island Research Alliance (RI Science & Technology Advisory Council), *Getting to the Bottom of Narragansett Bay Health: Single-Molecule Diagnosis of Microzooplankton in Response to Marine Conditions*. \$80k (40k to JRD); Chris Reid, Bryant (PI); JRD (co-PI). **Trainee Supervision**

Postdoctoral researchers

Dr. Teresa Mako (1/2020-9/2020) Paper-based diagnostics; Dr. Buddini Karawdeniya (5/2018-9/2018)

Graduate students

Elaine Foun (2010-2013 M.Sc., now at Fujifilm), Julie Whelan (2017 Ph.D.), Nuwan Bandara (Ph.D. Analytical Chemistry 2018, PDF Southern Methodist University Mechanical Engineering), Buddini Karawdeniya (Ph.D. Analytical Chemistry 2018, PDF Southern Methodist University Mechanical Engineering), Jon W. Nichols (M.Sc. Analytical Chemistry 2018, Nova Biomedical), Robert Chevalier

(Ph.D. Analytical Chemistry expected 2021), James Hagan (Ph.D. Analytical Chemistry expected 2022), Brian Sheetz (Ph.D. Analytical Chemistry expected 2023).

Award-winning graduate research

James Hagan: 2021 Eastern Analytical Symposium graduate research prize

Nuwan Bandara: 2016 Eastern Analytical Symposium research prize; 2015 URI Graduate School research fellowship.

Buddini Karawdeniya: 2017 Eastern Analytical Symposium research prize; 2016 URI Graduate School research fellowship, 2015 SciX conference poster prize.

Undergraduate students

1. Travis Leffert (B.Sc. 2010, staff scientist at Two Pore Guys, Inc.), 2. A.J. Laperche, 3. Cameron Frament (2009-2012; accepted to PhD Biophysics program at U. Colorado, Boulder), 4. Leslie McCabe (scientist with Blount Fine Foods), 5. Lucas Ginsberg (2012-2012; accepted to Berkeley chemistry PhD program), 6. Elana Viola, 7. Ian Tompkins (CHM 353, spring 2012; research assistant summer 2012), 8. Dan Wilson (CHM 353, summer 2012; PhD student Tufts), 9. Caitlin Ross (res. asst. summer 2012, 2013), 10. Josh Morimoto (2013), 11. Sarah Golden (2013, 2014), 12. Cory Lawton (2013), 13. Joshua Doyle (summer 2014), 14. Caitlin Masterson (2014, employed also at Hanna Instruments, now PhD student Brown Chemistry), 15. Jessika da Rocha Silva (2014; exchange student from Brazil), 16. Jayce Napolitano (2014), 17. Catherine Linh (2015 spring & summer), 18. Ben Rickson (2016 & 2017 summer), 19. Adriana Mendieta (2016 summer), 20. Michael Auten (2016-2017 academic), 21. Melissa Morris (2017 fall, 2018 fall), 22. Sam Toppa (2017 fall).

Award-winning undergraduate research:

Caitlin M. Masterson: 2014 Eastern Analytical Symposium research prize.

Cameron M. Frament: 2011 URI Undergraduate Research Initiative grant, 2012 Life Sciences, Physical Sciences and Engineering undergraduate student research award and 2012 Eastern Analytical Symposium undergraduate research prize (<http://www.uri.edu/news/releases/index.php?id=6440>).

Ian Tompkins: 2012 URI Undergraduate Research Initiative, *Research in 3D-Printing and rapid prototyping of high-sensitivity, low-cost diagnostic medical devices.*

Dan Wilson: 2013 URI Undergraduate Research Initiative, *How small is small? Measuring Bespoke Nanoscale Devices with a Real-time Visible Spectrometer.*

High School Students

Diondra Perillo (Bayview Academy junior & science intern, Spring 2013; majored in bioinformatics at U. Michigan), *Nanopore Current Blockage Simulations*, Nathan Meehan (Bishop Hendricken High School, Summer 2015), Priyanka Bonifaz (Spring 2016 term, Barrington High School junior; RI Science Fair top 10 plus: Yale Science and Engineering Award, American Chemical Society Award, Ricoh Sustainable Development Award); Blaine Lynch-Gadaleta (Bayview Academy junior & science intern, Spring 2018); Anna Khabaeva (ACS Project SEED Scholar, Summer 2018).

Teaching Experience

Fall 2021	<i>CHM 506: Chemical Analysis</i> <i>CHM 642: Graduate Chemistry Seminar</i>
Spring 2021	<i>CHM 644: Graduate Chemistry Seminar</i> <i>CHM 512: Advanced Analytical Chemistry II</i> <i>CHM 414: Instrumental Methods of Analysis (Laboratory; co with Prof. Jay Kim)</i>
Fall 2020	<i>CHM 506: Chemical Analysis</i> <i>CHM 642: Graduate Chemistry Seminar</i> <i>CHM 644: Graduate Chemistry Seminar</i>
Spring 2020	<i>CHM 412 & 414: Instrumental Methods of Analysis (Lecture & Laboratory)</i> -Pivoted lecture to online delivery in response to COVID-19 restrictions; -Developed new laboratory experiments to teach critical skills (e.g. electronics and data analysis, microscopy for material science) often lacking in chemistry students; -Developed an entirely new set of virtual experiments in response to COVID-19 restrictions (March-end of term).
Fall 2019	<i>CHM 506: Chemical Analysis</i> -Specially designed course for incoming graduate students to give an interdisciplinary overview of chemistry through chemical analysis. Course integrates textbook, literature, and discussion through a flipped classroom approach. <i>CHM 642: Graduate Chemistry Seminar</i> <i>CHM 644: Graduate Chemistry Seminar</i>
Spring 2019	<i>SABBATICAL</i>
Fall 2018	<i>CHM 506: Chemical Analysis.</i> <i>CHM 644: Graduate Chemistry Seminar</i>
Spring 2017	<i>CHM 414: Instrumental Methods of Analysis</i>
Fall 2016	<i>CHM 506: Chemical Analysis.</i> -Specially designed course for incoming graduate students to give an interdisciplinary overview of chemistry through chemical analysis. Course integrates textbook, literature, and discussion through a flipped classroom approach.
Spring 2015	<i>CHM 414: Instrumental Methods of Analysis</i> -Designed the lecture-based course to provide support to student self-instruction from sources including textbooks and the modern literature: the emphasis in this course was to prepare senior students for the experience of learning in a professional scientific setting outside of a university classroom.
Fall 2014	<i>CHM 506: Chemical Analysis.</i> -Specially designed course for incoming graduate students to give an interdisciplinary overview of chemistry through chemical analysis. Course integrates textbook, literature, and discussion through a flipped classroom approach.
Spring 2014	<i>CHM512: Advanced Analytical Chemistry II.</i> -Designed the course to fully integrate Mathematica symbolic and numeric mathematics into the study of analytical chemistry. Addressed a student unfamiliarity with computer programming skills and provided a rigorous framework for introducing new subjects.
Fall 2013	<i>CHM 511: Advanced Analytical Chemistry I.</i> -Delivered as a fully flipped classroom.
Spring 2010/11/12	<i>CHM 512: Advanced Analytical Chemistry II. (Graduate lecture course)</i> -Designed lecture course to emphasize problems from modern literature. -Incorporated a major independent project to design a new diagnostic

Fall 2009/10/11/12	instrument, and to consider commercial requirements of such a device. <i>CHM 642, 643, 644: Graduate Chemistry Seminars</i> <i>CHM 212: Quantitative Analysis (Undergraduate lecture + lab course)</i> -Delivered lectures that incorporated practical examples of principles from the modern chemical literature. -Designed and oversaw laboratory instruction.
Fall-Spring, Summer 2009/10/11/12	<i>CHM 353, 354: Undergraduate Student Research</i> -Designed independent research projects to introduce undergraduate students to modern chemical research.
Spring 2012	<i>CHM 642, 643, 644: Graduate Seminar Series</i> -Closely mentored students in writing abstracts, planning and delivering talks based on the literature and on their Ph.D. research.
Spring 2011-2014, 2016, 2017	<i>NUR 160: Exploring Global Health</i> guest lecture on <i>Portable Medical Diagnostics</i> .

Service

University of Rhode Island

- Committees & Commissions
 - Arts and Sciences Dean's Advisory Committee 2020-2023.
 - Arts and Sciences Committee on Research, Scholarship, and Creative Work 2020-2023.
 - Research Advisory Council Member, Office of VP Research & Development, URI, 2018-2022 (committee replaced by assoc. dean advisory council). To provide feedback to the VP R&D on research policies and activities at URI. Meetings called as-needed, ~1-2 times per semester. Formed as a less procedurally constrained alternative to the Senate-appointed Council for Research.
 - Founding member, Advisory Committee for RI Consortium for Nanoscience & Nanotechnology (2018-). In-person and online meetings 1-2 times per semester to assist director in strategic planning, proposal development, and career development.
 - Member of Hiring Committee for the RI Consortium for Nanoscience and Nanotechnology Director of Operations, Dr. Irene Andreu.
 - Faculty Appointee, Strategic Review of URI Research Foundation (Michael Katz), 2018 – Proposed ideas and advocated for how the Research Foundation could simultaneously benefit URI and faculty researchers. Part of the working group that had a number of in-person strategy sessions supported by off-line independent efforts. This effort informed the final rebranding of URI Research Foundation as URI Ventures.
 - URI President's Commission on People with Disabilities (2016-2019). Monthly meetings to advance the cause of access and accommodation at URI. My most profound contribution seems to have been to always question the acceptance of the *status quo*, to never accept "it's expensive" as a reason not to propose a solution, and to push for funded institutional-level approaches instead of small initiatives done as service.
 - University Laboratory and Chemical Safety Committee (2016-). To serve as a faculty voice in establishing umbrella safety procedures for the entire university.
 - Council for Research, appointed 2014-2017. Monthly meetings (~2 hours) to provide faculty input and oversight for research activities at the university. Evaluation of proposal development and career enhancement proposals over ~1 week on paper and 1-2 days in person each year.
 - Intellectual Property Committee 2015-2017 (seconded by Council for Research). Monthly ~2 hour meetings to evaluate the likelihood of invention disclosures to be patentable and market-competitive.
 - Met with prospective candidates in 2019 for the URI Licensing Associate position.
 - Champlin Foundations URI Provost's Proposal Review & Development Committee (summer 2015). After several years of being part of successful proposal teams, was asked

- to serve on the evaluation committee. During the course of providing constructive criticism, it became clear that I could help to build a more competitive applicant team and so I switched roles.
- Hiring Committee, Executive Director of University of Rhode Island American Association of University Professors (URI AAUP). Was interested in helping to develop a forward-looking ethos at a critical juncture for the union, with an emphasis on diversity and how faculty and institutional gains could be mutually beneficial.
 - Mentorship
 - Facilitator, URI's Metcalf Institute's Career Development Program Winter Intensive: How to Develop Effective Mentoring Relationships (January 28, 2022).
 - Facilitator, *How not to get lost as a cog in major projects: The challenges of team science* (October 29, 2021), Rhode Island EPSCoR C-AIM annual meeting.
 - Organized RI Summer Nano Talks student seminar series (2012, 2013). *Selected and coached students in an effort to build a nanoscience community at URI, and to help students build their presentation portfolio and skillset.*
 - Teaching
 - Guest lecturer for URI CSC 592 – No Boundary Thinking (Dec. 2018/Prof. Peckham)
 - Guest lecturer for URI NUR 160: Exploring Global Health: *Portable Medical Diagnostics*. March 30, 2011. April 4, 2012. April 3, 2013. April 2, 2014. April 6, 2016. April 6, 2017. (Prof. Schwartz-Barcott).
 - Invited lecturer for 2011 URI Honors Colloquium series
 - Guest lecturer for 2011 URI Honors class, Oct. 20, 2011
 - Chemistry building
 - Media outreach campaign *Essential 2 Healthcare: Chemistry Professor Jason Dwyer*. Print interview and photo campaign. Polling station information hand-outs.
 - Laboratory planning sessions. Proposed “showcase laboratory” concept to tie in with fundraising, proposed fully-enclosed working areas to ensure forward-compliance with health and safety regulations.
 - Academic Planning and Administrative
 - Graduate School Retreat planning session, Nov. 18, 2011.
 - Dean's External Advisory Council Presentation, June 19, 2013
 - Reviewed workplace health and safety documentation for maintenance and facilities personnel for Stacey Snow, URI Chemical Hygiene Officer.
 - Academic
 - Committee or Ph.D. Comprehensive Committee member: Santosha Ammu (PHYS, CHAIR 2011), Sreekanth Suravajjala (CHM, 2009), Glen Holden (CHM, 2010), Alexander Karabadzak (physics, 2011), Kaoru Tominaga (Pharmacy, 2013), Pratik Sheth (Pharmacy, 2013), Aihong Xi (ChemEng, 2012), Nicole Cook (CHM, 2013), Hanno Teiwes (MechEng, 2014), Austin Brown (CHM, 2016), Kurt Fastnacht (CHM, 2018), Timo Kuester (M.Sc. ChemEng, 2019), Zachary Brown (CHM, 2018), Benjamin Smith (M.Sc. CHM, 2017), Oleg Kazakov (CHM, 2018), Teresa Mako (CHM, 2019), Satu Heiskanen (CHM, 2019).

Department of Chemistry

- Committees
 - Chair, Graduate Curriculum Committee (2019-): Overseeing administration of graduate student curriculum progression, organizing qualifying examinations, counseling incoming graduate students, student advising, assessment, administering departmental awards. Running the welcome-to-URI Chemistry graduate student research session. COVID-19: Started “Unmask your research”, a graduate student research seminar, in close collaboration with the graduate student seminar committee to meet their vision and to better connect the incoming class to the department.
 - Chair, 2019 Physical/Analytical Hiring Committee. Ensuring fairness of search, adherence to diversity requirements, and striving to choose a candidate whose success

- will benefit the research, service, and teaching mission of URI. Administration of all reporting and 3-5 on-site visits.
- Graduate admissions committee (2009-2015). Detailed evaluation of applicant packages to ensure students were prepared for graduate studies, could contribute to the research and teaching missions of the university, and had skillsets and interests aligned with individual research group openings.
 - Graduate curriculum committee (2012-present): (2013-2014) Undertook a substantial overhaul of the graduate chemistry curriculum to institute greater interdisciplinary training. Mission statement: *To enable our students to make new discoveries and invent new applications, we must inculcate them with a rigorous understanding of fundamental chemical principles, delivered through courses firmly rooted in the interdisciplinary nature of the field.*
 - Search Committee for a Chemistry Lecturer (summer 2014)
 - Search Committee for a Physical/Analytical Chemist (2014)
 - Chair's Advisory Committee (2015-)
 - Chair, Chemistry Safety Committee (2015-2018)
 - Search Committee for a Physical/Analytical Chemist (2015) [two open positions]
 - Search Committee for a Physical/Analytical Chemist (2016)
 - Initiatives and Special Activities
 - A&S Events Fund (\$1000) to develop "URI 501" Graduate Student Welcome and Ongoing Guidance Sessions (Fall 2019). Conceived of and will coordinate 2-3 1-2 hour meetings in fall 2019.
 - Delivered a CV workshop to URI chemistry undergraduates, arranged by the departmental student organization (2/22/2017)
 - Spearheaded an annual chemistry research poster session for incoming and current chemistry graduate students (2013).
 - Instituted a number of departmental health and safety procedures including a no-glove policy in hallways and an improvement to the after-hours gas cylinder exchange procedure (2009-).
 - "Getting the most out of graduate school" event (April 26, 2012; August 9, 2013): an ongoing series of events to improve the chemistry graduate experience. Included participation from chemical engineering and mechanical engineering.
 - "Doors Open Chemistry" event (Jan. 19, 2012, Nov. 6, 2012, fall semester 2013-2015): a series of events to nurture an open, active and interdisciplinary research environment amongst student researchers.
 - Instituted Hypercube Scholar Program in the department (2011).
 - Introduced electronic scheduling for seminar speakers (2010-2011).
 - Initiated Dell printer program, leading to several new free advanced printers for department use (2009).
 - Arranged for URI students to be able to attend invitation-only (top schools) Novartis *Women in Chemistry* event taking place in parallel with the 2010 Boston ACS meeting.
 - Speakers hosted
 - 2019 – Clay Bennett (Tufts), Ben Saute (Telops—a company), James Reuther (UMass Lowell, Chemistry)
 - 2018 – Prof. Jing Zhao (UConn Chemistry), Prof. Ou Chen (Brown Chemistry)
 - 2017 – Prof. James Whitten (UMass Lowell Chemistry), Prof. Elizabeth Landis (College of the Holy Cross Chemistry).
 - 2016 – Dr. Erik T.J. Nibbering (Max Born Institute, Germany), Prof. Chia-Kuang (Frank) Tsung (Boston College Chemistry), Kwok-Fan Chow (UMass Lowell Chemistry), Min Chen (UMass Amherst Chemistry).
 - 2015 – Prof. Charles Mace (Tufts Chemistry).
 - 2014 – Prof. Charles Mace (Tufts Chemistry; cancelled due to weather), Prof. Ron Grimm (WPI Chemistry), Prof. Arne Gericke (WPI Chemistry),
 - 2013 – Sunshine Menezes (Metcalf Institute, URI Graduate School of Oceanography), Jianmin Gao (Boston College Chemistry), Mingdi Yang (UMass Lowell, Chemistry), Paul Whitford (Northeastern Physics), Wesley Wong (Biological Chemistry & Molecular

- Pharmacology and Pediatrics, Harvard), Xin Chen (Boston University, Chemistry), John Oliver (NabSys, Inc.), Prof. Robert Dempski (WPI, Chemistry),
- 2012- Ryan Hili (Harvard University Chemistry), Marco Masia (Boston University, Sassari)
- 2011-Sam Thomas III (Tufts Chemistry), Yong Wang (UConn, Engineering), Tse-Ming (Bob) Hsin (UMass Lowell Chemistry), Ethan Schonbrun (Rowland Institute, Harvard University)..
- 2010-Ludovico Cademartiri (Harvard University Chemistry), James Whitten (UMass Lowell Chemistry).
- 2009-Matthew Holden (UMass Amherst Chemistry), Murugappan Muthukumar (UMass Amherst Polymer Science and Engineering), Andre Marziali (UBC Canada Physics).

Professional (External) Service

- Panelist, *Forming cross-disciplinary collaborations*. NIH NHGRI Advanced Genomic Technology Development Meeting. Virtual meeting, May 27-29, 2021.
- Local Conference Chair, SciX 2021, Providence, RI, September 26-Oct. 1, 2021.
- NSF CMI Panelist, 2020.
- NSF AccelNet *ad hoc* reviewer, 2020.
- Panelist, National Academies of Sciences, Engineering, and Medicine, NRC Research Associateship Program, 2019. By invitation (Mark Hayes, ASU), 2 days on-site at Arnold and Mabel Beckman Center of the National Academies of Sciences and Engineering (Irvine, CA), followed by 3 online review sessions.
- Councilor, AES Electrophoresis Society (2018-2021). Monthly teleconferences & annual board meeting: society strategic planning, financial management, conference planning, outreach.
 - Awards Chair, 2018-2020. Administering three annual awards from competition to conference awards sessions – AES Distinguished Service Award, AES Lifetime Achievement Award, and AES Mid-Career Award.
- Session Chair, AES Mid-Career Award Session, Annual AES Meeting at SciX 2018, Atlanta, GA, October 21-24. Designing program, inviting speakers and chairing session.
- Co-chair of AES Symposium, FACSS SciX 2016 & 2017. Planning a 6-session research symposium each year by choosing the focus and session chairs, and overseeing session organization.
- Symposium Session Chair, FACSS SciX 2015 *Biopolymers in Electric Fields*, Providence, RI, Sept. 27-Oct. 2, 2015.
- ACS Symposium Presider: *Nanotechnology for Analytical Sensing and Spectroscopy Based Applications*, ACS Fall National Meeting, Boston, USA, August 16-20, 2015.
- ACS Session Chair, Spring National Meeting, New Orleans 2013, *Transformative Nanotechnologies: Energy and Environment, Solutions and Challenges (Focused on Materials Development and Analyses)*
- Advisory Board, 3rd Next Generation Sequencing Conference, Global Technology Community, Monrovia, CA (2012-2013)
- MITACS Accelerate Proposal Reviewer (Canada) 2020
- NIH Grant Review Panel Member: NHGRI Revolutionary Sequencing Technology grants (2/23/12, 3/26/14; 3/2016; 12/2018; 11/2020). This funding program was glowingly featured in Nature (2014) **507**, 273 and 294-295—a rare feat for a granting program.
- NSF external grant reviewer (2009, 2012, 2013)
- Reviewer

<ul style="list-style-type: none"> ○ ACS Nano ○ ACS Sensors ○ Analytical Chemistry ○ Analyst ○ Cell Biochemistry and Biophysics ○ ELECTROPHORESIS ○ JACS (4 times) 	<ul style="list-style-type: none"> ○ Journal of Chemical Physics ○ Journal of Physics: Conference Series ○ Journal of Physical Chemistry (3 times) ○ Journal of Physical Chemistry Letters (2015)
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- Journal of Photochemistry & Photobiology, B: Biology
 - Langmuir
 - MRS Proceedings
 - Nano Letters
 - Nanoscale
 - Nature Biotechnology
 - Nature Nanotechnology (4 times)
 - PLoS One
 - Physical Chemistry Chemical Physics (2 times)
 - Sequencing
 - U.S. Environmental Protection Agency (EPA) Prize Competition - 2017 Innocentive Advanced Septic System Nitrogen Sensor Challenge
- External reviewer (in French), Ph.D. Chemistry committee, Alexandra Aubé, University of Montreal, Canada
 - Natural Sciences Engineering and Research Council of Canada (NSERC) Collaborative Research and Development (CRD) site visit external examiner for a joint academia-industry research proposal (October 2018). Preliminary evaluation of proposal, 1-day site visit and follow-up report covering issues from science, commercialization potential, and diversity.
 - ACS Session Chair & Organizer, Fall National Meeting, Boston, 2010, *Clinical Applications of Sensors*.
 - Session Chair, Nanopores Conference 2012, Lanzarote, Spain.
 - Session Chair, GTC 2nd Next Generation Sequencing Conference, Boston, USA, May 30-31, 2012
 - Policy pre-2009
 - DFAIT Canada-Germany science panel discussion on scientific cooperation, Photonics Research Ontario round-table on scientific cooperation, lab tour for Ontario Asst. Deputy Minister of Technology, femtosecond laser facility planning session for the Canadian Light Source.

Public Outreach and Media

- 2020 *To See the Sea or Not to See: Optical and Nonoptical Sensors for Marine Environments*. Flash talk, The Blue Innovation Symposium: Sensors and the Next Wave of Data, Newport, RI, January 14-16, 2020.
- 2019 **Ocean State Innovators, Boston Globe**, Dec. 23, 2019. “Ask questions, especially when everybody seems to accept the way something is done”.
- 2019 Sensor Technology Development for Ecosystem and Economic Impact: A C-AIM Snapshot. Provided a scientific overview of NSF C-AIM program (NSF OIA-1655221) to the RI Innovation Community gathered in Providence for a meeting on “The Blue Economy”: Bluetech Connect at Venture Café, Providence. Nov. 7, 2019. Providence, RI.
- 2019 *Developing Nanopore Technology for Medical Diagnostics*. Research Profile, URI Research Momentum Magazine.
- 2019 Session Leader - RI C-AIM Career Development Program Science Communication Summer Intensive. *Presenting and Pitching Your Expertise and Patent Ideas*. URI, July 12, 2019. Also proposed the focus on commercialization and career training, including a July 11 session: *Primer on Technology Commercialization and Intellectual Property*. Initiated and planned session with Dr. Sunshine Menezes, Metcalfe Institute, URI.
- 2019 Tech Talk – Sensor Development: Coastal Ecology Assessment, Innovation, and Modeling. The Southeastern New England Defense Industry Alliance, Charlestown, RI, USA, Feb. 14, 2019. Business networking session with a vital New England industry group, organized through RI EPSCoR C-AIM and URI’s Business Engagement Center.
- 2019 Highlighting High-Profile URI Research Mentions on URI Public Outreach Platforms e.g. <https://chemistrycommunity.nature.com/users/172892-buddini-karawdeniya> Active engagement in crafting outreach stories, e.g. for URI Research Momentum (in preparation)
- 2019 ACS Project SEED Student. Supervised summer research through Project ACS SEED, an 8-10 week long national program to help economically disadvantaged high school students by letting

- them work in real laboratories, with real scientists serving as their mentors. The student is now in health studies at URI.
- 2017 Guest session leader at April 2017 Dreyfus Foundation sponsored weeklong science camp for middle school girls. Prepared 2-3 demonstrations for an hour-long interesting and inspiring talk on science.
- 2014 •Invited lecture, *Science Communication* at the Sustainable Nanotechnology Organization-sponsored *Communicating Nanostuff* Workshop at the 3rd USA Science and Engineering Festival, Washington, D.C., April 25, 2014.
•Guest session leader at April 2014 Dreyfus Foundation sponsored weeklong science camp for middle school girls. Prepared 2-3 demonstrations for an hour-long interesting and inspiring talk on science.
- 2013 •Guest lecture, *Nanoscience and Nanotechnology*. Osher Lifetime Learning Institute *The Cutting Edge: Thinking Big in Scientific Inquiry at URI* course, Kingston, RI, November 6, 2013 with graduate students guest speakers.
•Guest lecture, *Nanofluidic cell for high-resolution, in-liquid transmission electron microscopy*. Osher Lifetime Learning Institute *The Future is Here* course, Kingston, RI, May 1, 2013.
•Faculty mentor (Ian Tompkins) for a guest lecture on 3D printing at the Osher Lifetime Learning Institute, Kingston, RI, May 8, 2013.
•Laboratory tour for Cranston Outreach Math Program (August 14, 2013)
•Guest session leader at April 2013 Dreyfus Foundation sponsored weeklong science camp for middle school girls. Prepared 2-3 demonstrations for an hour-long interesting and inspiring talk on science.
•Mentor for high school student Diondra Perillo (Bayview Academy, Spring 2013), *Nanopore Current Blockage Simulations*
- 2012 •*Warwick Beacon*, quoted in “It’s not rocket science” and “Students display questioning minds at science fair”, March 20, 2012.
•*Providence Phoenix Q&A on nanotechnology* 9/25/11
•*Front page media coverage of NSF CAREER award in Sun Publishing Co. newspapers* (Wood River Press, Charlestown Press, etc.)
•*Coverage of NSF CAREER award in Providence Business News*
- 2010-2013 *Rhode Island Science & Engineering Fair* Judge.
- 2010 *Essential 2 Healthcare: Chemistry Professor Jason Dwyer*
<http://essential2ri.org/YesOn2/essential-2/essential-2-healthcare-chemistry-professor-jason-dwyer/>
Photo and text-based outreach for URI Chemistry building campaign explaining my research focus on rapid portable diagnostics. Numerous written drafts to ensure the need for a new building was made without denigrating the quality of work done currently.
- 2008 •*Making the MOST out of media relations*
Department of Physics, University of British Columbia.
Conceived of and organized a presentation by the public affairs office at UBC to a general audience of scientists on why and how to talk to the media about science. Subsequently organized a similar media training presentation for the School of Population and Public Health (2008)
•*Rising Stars of Research*
Poster competition judge for Canadian competition at the University of British Columbia.
- 2007 *UN Migrant Workers’ Day*
Rundfunk Berlin-Brandenburg (RBB) Radio
Interview with a German radio news magazine about my experience as a foreign scientific worker in Germany.
- 2007 *Ultrafast laser science*
Hauptstadtstudio Berlin, ZDF Morgenmagazin
Interview with a German television news magazine focusing on ultrafast science and technology in Berlin. Provided visual demonstrations of femtosecond laser operation.
- 2004 *One Second/Bullet Time*

- Discovery Channel/France 5
Documentary feature interview (20 minutes of 60 minutes total length) discussing what happens on the timescale of femtoseconds, how it is possible to investigate these ultrafast processes and why it is interesting to know about them. Several days were spent offscreen providing technical explanations.
- 2003 *Femtosecond electron diffraction*
SpaceTV news (Canadian national television network)
Media interview explaining the basics of the technique of femtosecond electron diffraction for a general audience, as well explaining what insights the technique offers into the physical world.
- 2001 *Frontiers in ultrafast science*
Organized an introductory lecture for undergraduate students at Trinity College, Canada, on recent advances in ultrafast optical science.
- 2000 *Why science?*
Organized a gathering of undergraduate science students and research leaders in chemistry, physics and mathematics with the goal of inspiring the next generation of scientists.

Entrepreneurship & Consulting

- Co-founder: Insight Nanofluidics
- Awarded 2012 Ontario Centres of Excellence *Market Readiness* Phase I grant, \$25k; 2013 Ontario Centres of Excellence Market Readiness Phase 2A, \$125k, VentureStart \$30k.

Special Training

- 2021 *National Research Mentoring Network for a Diverse Biomedical Workforce (NRMN) & Center for the Improvement of Mentored Experiences in Research. Entering Mentoring Mentor Training Workshop.*
Facilitated by: Samantha Meenach, Ph.D., Bryan Dewsbury, Ph.D., Aisling Caffrey, Ph.D., M.S., and Michelle Fontes, M.A. Including Effective Communication, Promoting Professional Development, Aligning Expectations, Addressing Equity and Inclusion, and Articulating Your Mentoring Philosophy and Plan.
URI Inclusive Mentoring Mentor Training Workshop.
Facilitated by: Samantha Meenach, Ph.D., Bryan Dewsbury, Ph.D., Aisling Caffrey, Ph.D., M.S., and Michelle Fontes, M.A. Including Knowing and Understanding Your Why, Chance at Birth, Power and Privileges in the Research Sciences, and Scenario-Based Mentoring.f
- 2008 *Molecular Biology Techniques Workshop*
Dr. David Ng, Michael Smith Labs, University of British Columbia.
-week-long hands-on training in molecular biology techniques.
- 2008 *Laboratory Biological Safety*
University of British Columbia, Dept. of Health, Safety and Environment.
National Coaching Certification Program, Levels 1 & 2 Theory
Coaching Association of Canada. Formal training in coaching approaches.

Professional Memberships

- Member, American Chemical Society (2009-)